Novel Photodegradation of the Antifungal Antibiotic Pyrrolnitrin in Anhydrous and Aqueous Aprotic Solvents

Magoichi Sako,*^{,†} Toshiyuki Kihara,[†] Mihoko Tanisaki,[†] Yoshifumi Maki,[†] Akira Miyamae,[‡] Toshio Azuma,[‡] Shigetaka Kohda,[‡] and Takashi Masugi[‡]

Laboratory of Medicinal Chemistry, Gifu Pharmaceutical University, Mitahora-higashi, Gifu 502-8585, Japan, and Analytical Research Laboratories, Fujisawa Pharmaceutical Co., Ltd., Kashima, Yodogawa, Osaka 532-8514, Japan

sako@gifu-pu.ac.jp

Received June 18, 2001

The UV irradiation of pyrrolnitrin (1a), which is an antibiotic clinically useful against dermatophytosis and possesses a unique 2-(pyrrol-3-yl)nitrobenzene moiety in the molecule, in an anhydrous aprotic solvent resulted in the exclusive formation of transient 7,4'-dichlorospiro[1,3-dihydrobenzo-(c)isoxazole-3,3'-pyrrolin-2'-one] (2a) via the intramolecular oxidation of the juxtaposed pyrrole ring by the triplet-excited nitro group. The irradiation in an aqueous aprotic solvent, however, allowed the concurrent occurrence of intramolecular cyclization by the singlet-excited nitro group in 1a and the hydroxylation at the 2-position of the pyrrole ring by water to afford 3,7-dichloro-8-hydroxy-8,8a-dihydropyrrolo[2,3-b]indol-2-one (3a), accompanied by the formation of 2a. Elongation of the irradiation time in these photoreactions caused a rapid consumption of the products, 2a and 3a, to give undetermined polar polymeric products. The present results indicate that the photodegradation of **1a** is significantly influenced by the presence of water in the reaction media and by the nature of its excited state. Thus, the loss of the antifungal activities by the photosensitive antibiotic 1a was chemically proved.

I. Introduction

Pyrrolnitrin (1a), 3-chloro-4-(3-chloro-2-nitrophenyl)pyrrole,¹ is an antibiotic produced from L-tryptophan² by Pseudomonas species,³ Burkholderia cepacia,⁴ Myxococcus fulvus,⁵ Serratia species,⁶ and Enterobacter ag*glomerans*.⁷ The antibiotic **1a** is on the market today as an externally used chemotherapeutic agent for tinea pedis and other types of dermatophytosis⁸ on the basis

(3) a) For P. pyrrocinia: Imanaka, H.; Kousaka, M.; Tamura, G.; Arima, K. J. Antibiot. (Tokyo), Ser. A 1965, 18, 205-206. (b) For P. Aureofaciens, P. fluorescens, and P. multivorans: Keller, S.; Wage, T.; Hohaus, K.; Holzer, M.; Eichhorn, E.; van Pee, K.-H. Angew. Chem., Int. Ed. 2000, 39, 2300–2302. Corbell, N.; Loper, J. E. J. Bacteriol. **1995**, *177*, 6230–6236. Elander, R. P.; Mabe, J. A.; Hamill, R. H.; Gorman, M. *Appl. Microbiol.* **1968**, *16*, 753–758. Lively, D. H.; Gorman, M.; Haney, M. E.; Mabe, J. A. Antimicrob. Agents Chemother. 1966, 462-469. (c) For P. acidula: Ajisaka, M.; Jomon, K.; Kubochi, Y. JP 462–469. (c) For *P. acidula*: Ajisaka, M.; Jomon, K.; Kubochi, Y. JP 7222797 1969; *Chem. Abstr.* **1972**, *77*, 99667x. (d) For *P. cepacia*: Roitman, J. N.; Mahoney, N. E.; Janisiewicz, W. J. *Appl. Microbiol. Biotechnol.* **1990**, *34*, 381–386. Homma, Y.; Sato, Z.; Hirayama, F.; Konno, K.; Shirahama, H.; Suzui, T. *Soil Biol. Biochem.* **1989**, *21*, 723– 728. Kuo, M. H.; Hsu, Y. H.; Lo, M. C.; Soong, T. S. *Chung Yang Yen Chiu Yuan Chih Wu Yen Chiu So Chuan Kan* **1989**, *9*, *245*–262.

(4) El-Banna, N.; Winkelmann, G. J. Appl. Microbiol. 1998, 85, 69-78.

of its inhibitory activities toward NADH- and succinatesupported electron transport in mitochondoria^{4,9} and on the destructive influence to the cell membrane by combination with phospholipids.¹⁰ Furthermore, the antibiotic 1a has been recently documented to play a very important role as an effective biocontrol agent for the soilborne fungal pathogens in a variety of plants¹¹ and to exhibit an appreciable inhibiting activity against Mycobacterium tuberculosis and M. avium.¹²

This antibiotic has a unique structure; i.e., the dihedral angle of the pyrrole ring to the benzene ring was shown to be 52° and the plane through the nitro group is almost at a right angle (88°) to the plane of the benzene ring on the bais of its X-ray crystal analysis,¹³ suggesting the absence of a long conjugation system in the molecule. An extreme lability of **1a** [λ_{max} 252 (ϵ 7.5 \times 10³) nm, accompanied by the tailing of the peak to 400 nm] upon exposure to sunlight, however, was observed in early investigations, resulting in significant destruction of its antifungal activity.¹⁴ In view of these observations and from our (M.S. and Y.M.) continuous interests in the

[†] Gifu Pharmaceutical University.

[‡] Fujisawa Pharmaceutical Co., Ľtd.

⁽¹⁾ Imanaka, H.; Kousaka, M.; Tamura, G.; Arima, K. J. Antibiot. (Tokyo), Ser. A **1965**, 18, 207–210.

⁽²⁾ Van Pee, K.-H.; Ligon, J. M. Nat. Prod. Rep. 2000, 17, 157-164; Hammer, P. E.; Burd, W.; Hill, D. S.; Ligon, J. M.; van Pee, K.-H. FEMS Microbiol. Lett. **1999**, 180, 39–44; Dolfing, J. FEMS Microbiol. Lett. 1998, 167, 271-274; Hohaus, K.; Altmann, A.; Burd, W.; Fisher, I.; Hammer, P. E.; Hill, D. S.; Ligon, J. M.; van Pee, K.-H. Angew. Chem., Int. Ed. Engl. **1997**, *36*, 2012–2013 and references therein

⁽⁵⁾ Gerth, K.; Trowitzsch, W.; Wray, V.; Hoefle, G.; Irschik, H.; Reichenbach, H. *J. Antibiot.* **1982**, *35*, 1101–1103.

⁽⁶⁾ Kalbe, C.; Marten, P.; Berg, G. Microbiol. Res. 1996, 151, 432-439.

⁽⁷⁾ Chernin, L.; Brandis, A.; Ismailov, Z.; Chet, I. Curr. Microbiol. **1996**, *32*, 208–212.

⁽⁸⁾ Uchida, K.; Yamaguchi, H. Jpn. J. Antibiot. 1999, 52, 68-74. Tawara, S.; Matsumoto, S.; Watanabe, Y.; Hirose, T.; Matsumoto, Y.; Nakamoto, S.; Kamimura, T.; Yamaguchi, H. Shinkin to Shinkinsho 1989, 30, 266–272. Lu, Y.-C. Tai-Wan I Hsueh Hui Tsu Chih 1971, 70, 449–452; Gordee, R. S.; Matthews, T. R. Appl. Microbiol. 1969, 17, 690–694; Antimicrob. Agents Chemother. 1967, 378–387. Nishida, M.; Matsubara, T.; Watanabe, N. J. Antibiot. (Tokyo), Ser. A 1965, 18, 211-219.

⁽⁹⁾ Kawai, K.; Shiojiri, H.; Watanabe, R.; Nozawa, Y. Res. Commun. Chem. Pathol. Pharmacol. 1983, 39, 311-319. Warden, J. T.; Edwards, Cnem. Pathol. Pharmacol. 1983, 39, 311–319. Warden, J. T.; Edwards,
D. L. Eur. J. Biochem. 1976, 71, 411–418. Lambowitz, A. M.; Slayman,
C. W. J. Bacteriol. 1972, 112, 1020–1022. Wong, D. T.; Horng, J.-S.;
Gordee, R. S. J. Bacteriol. 1971, 106, 168–73. Wong, D. T.; Airall, J.
M. J. Antibiot. 1970, 23, 55–62. Tripathi, R. K.; Gottlieb, D. J.
Bacteriol. 1969, 100, 310–318.
(10) Nose, M.; Arima, K. J. Antibiot. (Tokyo) 1969, 22, 135–143.

photochemical reactivity of antibiotics and medicines,¹⁵ we attempted to clarify the photodegradation modes of **1a**.

II. Results and Discussion

Photoreactions of Pyrrolnitrin (1a). The antibiotic 1a was stable in dry acetonitrile even under reflux for 1 day. In sharp contrast to the high stability under thermal conditions, 1a was extremely labile upon exposure to UV (>280 nm) light resulting in a smooth consumption (Φ = 0.1) accompanied by color change in the solution from pale yellow to brown. The TLC analysis of the reaction mixture showed that a polar compound (2a) was formed as the major product during the initial stage, together with undetermined polar polymeric products, e.g., the yield of 2a was 28% after the irradiation for 10 min, though the 35% recovery of the starting 1a was observed under the conditions employed. Elongation of the irradiation time caused degradation of the photoproduct **2a** [λ_{max} 286 (ϵ 3.0 \times 10³) nm] leading to polymeric compounds of which the ¹H NMR spectrum showed the appearance of unassignable multiplet signals in the aromatic region. The use of dry THF, chloroform, or diethyl ether as a solvent for the photoreaction of 1a was more effective (47-72% yields after a 10-min irradiation) for the formation of 2a. Although 1a is thermally stable in 5% aqueous acetonitrile, the UV irradiation of 1a in the same aqueous solvent allowed the formation of another polar compound (3a) in 20% yield together with 2a (24%). The product

Scheme 1. Formation of Methanol Adduct 4 from



distribution of **2a** and **3a** in this photoreaction was significantly dependent on the water content in the reaction media; an increase in the water content of the solvent caused the preferential formation of **3a** than that of **2a**, e.g., the yields of **2a** and **3a** in 20% aqueous acetonitrile were 11% and 40%, respectively, after a 10 min irradiation.

Structures of the Photoproducts 2a and 3a. The product **2a** was very unstable even in an aprotic solvent and at ambient temperature and was converted into undetermined polar polymeric products.¹⁶ However, purification of the reaction mixture of **1a** by flash chromatography using a very short column and toluene–ethyl acetate as an eluent followed by recrystallization from toluene allowed the isolation of **2a**, mp 225 °C dec, as a colorless amorphous powder in the pure state. Contrary to the chemical property of **2a**, the product **3a** was a fairly stable even in a warmed protic solvent and recrystallized from ethanol to afford colorless prism crystals, mp 175 °C dec. Microanalytical results showed that the photoproducts **2a** and **3a** are formulated as C₁₀H₆N₂O₂Cl₂, which is the same as that of the starting **1a**.

The remarkable spectral changes of **2a** from **1a** are as follows: (a) a significant shift in the three proton signals assignable to the benzene-ring protons to the higher fields $[\delta_{\rm H} 7.57 \text{ (dd, } J = 2, 8 \text{ Hz}), 7.69 \text{ (t, } J = 8 \text{ Hz}), \text{ and } 7.75$ (dd, J = 2, 8 Hz) ppm for **1a**; $\delta_{\rm H}$ 7.03 (dd, J = 2, 7 Hz), 7.08 (t, J = 7 Hz), and 7.41 (dd, J = 2, 7 Hz) ppm for **2a**] in the ¹H NMR spectrum was observed, implying a drastic structural change in the nitro group of 1a during the reaction. (b) the disappearance of one proton signal for the pyrrole-ring from the aromatic region [$\delta_{\rm H}$ 6.9 and $\delta_{\rm H}$ 7.15 ppm for **1a**: $\delta_{\rm H}$ 7.28 (d, J = 2 Hz) ppm for **2a**] and appearance of a γ -lactam moiety [v 1733 cm⁻¹ ($\delta_{\rm C}$ 174.8 ppm) for the amide carbonyl group; v 3190 cm⁻ $(\delta_{\rm H} 10.5 \text{ ppm})$ for the amino group] in the ¹H NMR and IR spectra were observed, indicating the occurrence of the oxidation at the α -position of the pyrrole ring in **1a** during the photoreaction. On the basis of these spectral data, the structure of 2a was reasonably assigned as 7,4'dichlorospiro[1,3-dihydrobenz(c)isoxazole-3,3'-pyrrolin-2'one], which has a novel type of spiro heterocyclic ringsystem in the molecule. The structure having a spiroasymmetric carbon was well supported by its HMBC spectrum showing the presence of cross-peaks of the spiro-centered carbon [$\delta_{\rm C}$ 89.9 (C-3) ppm] to the benzenering proton [$\delta_{\rm H}$ 7.03 (H-4) ppm], the pyrrole NH proton $[\delta_{\rm H} 9.96 \text{ (br d, } J = 2 \text{ Hz}) \text{ ppm}]$, and the pyrrole-ring proton [$\delta_{\rm H}$ 7.28 (H-5'; d, J = 2 Hz) ppm]. The isolated product 2a was optically inactive. Further structural proof was obtained upon treatment of **2a** with methanol at room temperature leading to a ring-opening adduct

^{(11) (}a) For rice: Rosales, A. M.; Thomashow, L.; Cook, R. J.; Mew, T. W. *Phytopathology* **1995**, *85*, 1028–1032. (b) For melon, cucumber, tomato, onion, lily, kidney bean, Welsh onion, and adzuki bean: Sarniguet, A.; Kraus, J.; Henkels, M. D.; Muehlchen, A. M.; Loper, J. cotton seedling: Hill, D. S.; Stein, J. I.; Torkewitz, N. R.; Morse, A. M.; Howell, C. R.; Pachlatko, J. P.; Becker, J. O.; Ligon, J. M. Appl. Environ. Microbiol. **1994**, 60, 78–85; Howell, C. R.; Stipanovic, R. D. Phytopathology **1979**, 69, 480–482. (e) For wheat straw: Pfender, W. F.; Kraus, J.; Loper, J. E. Phytopathology, 1993, 83, 1223-1228. (f) For rose flower: Jayaswal, R. K.; Fernandez, M.; Upadhyay, R. S.; Visintin, L.; Kurz, M.; Webb, J.; Rinehart, K. *Curr. Microbiol.* **1993**, *26*, 17–22. Hammer, P. E.; Evensen, K. B.; Janisiewicz, W. J. Plant Dis. 1993, 77, 283-286. (g) For sunflower: McLoughlin, T. J.; Quinn, J. P.; Bettermann, A.; Bookland, R. *Appl. Environ. Microbiol.* **1992**, 58, 1760–1763. (h) For raspberry: Goulart, B. L.; Hammer, P. E.; Evensen, K. B.; Janisiewicz, W.; Takeda, F. J. Am. Soc. Hortic. Sci. 1992, 117, 265-270. (i) For apples and pears: Janisiewicz, W.; Yourman, L.; Roitman, J.; Mahoney, N. Plant Dis. 1991, 75, 490-494. (j) For strawberry: Takeda, F.; Janisiewicz, W. J.; Roitman, J.; Mahoney, N.; Abeles, F. B. Hort Science 1990, 25, 320-322. (k) For radish seedling: Homma, Y.; Suzui, T. Nippon Shokubutsu Byori Gakkaiho 1989, 55, 643-651. (l) For rapeseed seedling: Dahiya, J. S.; Woods, D. L.; Tewari, J. P. *Bot. Bull. Acad. Sin.* **1988**, *29*, 135– 142. (m) For natural products with antifungal activity from Pseudomonas bio-control bacteria: Ligon, J. M.; Hill, D. S.; Hammer, P. E.; Torkewitz, N. R.; Hoffman, D.; Kempf, H.-J.; van Pee, K.-H. Pest. Manage. Sci. 2000, 56, 688-695.

 ⁽¹²⁾ Di Santo, R.; Costi, R.; Artico, M.; Massa, S.; Lampis, G.;
 Deidda, D.; Pompei, R. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 2931–2936.
 (13) Morimoto, Y.; Hashimoto, M.; Hattori, K. *Tetrahedron Lett.* **1968**, 209–211.

⁽¹⁴⁾ Arima, K.; Imanaka, H.; Kousaka, M.; Fukuta, A.; Tamura, G. J. Antibiot. (Tokyo), Ser. A **1965**, *18*, 201–204.

⁽¹⁵⁾ Sako, M.; Takeda, Y.; Hirota, K.; Maki, Y. Heterocycles 1996,
42, 31-34; Sako, M.; Nagai, K.; Maki, Y. J. Chem. Soc., Chem. Commun. 1993, 750-751. Sako, M.; Oyabu, I.; Hirota, K.; Maki, Y. J. Chem. Soc. Chem. Commun. 1991, 601-602. Maki, Y.; Shimada, K.; Sako, M.; Kitade, Y.; Hirota, K. Chem. Pharm. Bull. 1988, 36, 1714-1720. Sako, M.; Shimada, K.; Hirota, K.; Maki, Y. Tetrahedron Lett. 1986, 27, 3877-3880. Maki, Y.; Sako, M. J. Chem. Soc., Chem. Commun. 1978, 836-838; J. Am. Chem. Soc. 1977, 99, 5091-5096.

⁽¹⁶⁾ For this reason, all attempts to prepare a single crystal of **2a** suitable for X-ray crystal analysis were unsuccessful.



Figure 1.



Figure 2. ORTEP drawing of the X-ray crystal structure of **3a**.

(4), mp 140 °C dec, of which the formation can be reasonably explained by considering the addition of methanol to the 5'-position and the cleavage of the C_3-O_2 bond in **2a** as shown in Scheme 1.

Characteristics of the spectra of the photoproduct 3a compared with those of 1a are as follows: (a) A slight signal-shift of the benzene-ring protons in 1a to the higher fields [$\delta_{\rm H}$ 7.18 (t, J = 8 Hz), 7.48 (dd, J = 3, 8Hz), and 7.57 (dd, J = 3, 8 Hz) ppm for **3a**] in the ¹H NMR spectrum was observed, suggesting a small structural change in the nitro group of **1a** during the reaction. (b) The disappearance of two proton signals for the pyrrole ring from the aromatic region and appearance of a signal attributed to a γ -lactam moiety [v 1720 cm⁻¹ ($\delta_{\rm C}$ 169.8 ppm) for the amide carbonyl group; v 3255 cm⁻¹ $(\delta_{\rm H} 9.15 \text{ ppm})$ for the amide N–H group] and a methine proton [$\delta_{\rm H}$ 5.89 (d, J = 3 Hz) ppm; $\delta_{\rm C}$ 87.0 ppm] in the NMR and IR spectra were observed. This is indicative of oxidation at the α -position on the pyrrole ring in **1a** and of the intramolecular cyclization at another α -position of the pyrrole ring. On the basis of these spectral data and its HMBC spectrum, the structure of 3a was assigned to 3,7-dichloro-8-hydroxy-8,8a-dihydropyrrolo[2,3blindol-2-one and was finally confirmed by its X-ray crystallographic analysis (see Figure 2). The product **3a**, which has a single chiral center [$\delta_{\rm C}$ 87.0 (C-8a) ppm] in the molecule, is a 1:1 mixture of stereoisomers from X-ray analysis.

Mechanistic Aspects. No interconversion between the photoproducts **2a** and **3a** was observed under analogous irradiation conditions, clearly indicating that these products are formed via different reaction paths during the present photoreaction. Further experiments were carried out to elucidate the reaction mechanisms for the formation of the spiro-benzoisoxazoline **2a** and the pyrroloindoline **3a** during the photoreaction of **1a** by employment of a triplet-sensitizer and intersystem-crossing accelerator. The consumption of **1a** and the formation of



Figure 3. Four samples of acetrophenone (0.2) mmol) in dry acetonitrile containing pyrrolitrin **1a** (0.2, 0.4, 0.6, and 0.8 mmol) were prepared. The fluorescence intensities of these samples obtained by excitation at 282 nm were measured at the emission wavelength of acetophenone (308 nm). If⁰: The fluorescence intensity of acetophenone itself. If: The fluorescence intensities of the samples containing **1a**.



Figure 4. Effects of triplet sensitizer on the Photoreactions of pyrrolnitrin **1a**.

2a during the photoreaction using the aqueous acetonitrile as a solvent were markedly accelerated by the addition of acetophenone, a well-known triplet sensitizer,¹⁷ in a concentration dependent manner as shown in Figure 4 (see also Figure 3). In contrast to these facts, the formation of 3a was evidently decreased under the triplet-sensitized conditions. The addition of tribromomethane, an accelerator for the intersystem-crossing from a singlet-excited state to a triplet-excited state,¹⁸ to the photoreaction media resulted in a significant increase in the formation of **2a** and the decrease in the formation of **3a** in an accelerator-concentration dependent way (see Figure 5). These facts strongly suggest that the triplet-excited state of **1a** is related to the formation of 2a and its singlet-excited state plays a role in the formation of 3a.

Taking the above facts and the photochemical reactivity of the nitro function^{19,20} into consideration, we propose a plausible reaction sequence for the formation of 2aduring the photoreaction of 1a involving the intramolecular addition of the triplet-excited nitro group to the

⁽¹⁷⁾ For examples, see: Ono, I.; Sato, S.; Fukuda, K.; Inayoshi, T. *Bull. Chem. Soc. Jpn.* **1997**, *70*, 2051–2055. Suyama, K.; Ito, K.; Tsunooka, M. J. Polym. Sci. Part A: Polym. Chem. **1996**, *34*, 2181–2187.

⁽¹⁸⁾ For an example, see Nicodem, D. E.; da Cunha, M. F. V. J. Photochem. Photobiol., A 1997, 107, 165–167.

⁽¹⁹⁾ Lye, J.; Freeman, H. S. Adv. Colour Sci. Technol. **1999**, *2*, 124–141. D'Auria, M.; Ferri, T.; Mauriello, G.; Pesce, A. Tetrahedron **1999**, *55*, 2013–2024. Fasani, E.; Mella, M.; Albini, A. J. Chem. Soc., Perkin Transl. *1* **1992**, 2689–2692. Mariano, P. S.; Stavinoha, J. L. In Synthetic Organic Photochemistry, Horspool, W. M., Ed.; Plenum Press: New York, 1984; pp 145–258. Morrison, H. A. In The Chemistry of the Nitro and Nitroso Groups; Feuer, H., Ed.; Interscience Publishers: New York, 1969; pp 165–213.



Figure 5. Effects of IC-accelerator on the photoreactions of pyrrolnitrin **1a**.





pyrrole ring (cf. **A**) followed by collapse to give 2a as shown in Scheme 2. An analogous type of reaction sequence in the photochemistry of nitro compounds has been generally accepted.²⁰

The formation of **3a**, involving hydroxylation at the 2-position of the pyrrole ring in **1a**, is not reasonably explained by virtue of the mechanism proposed for the formation of **2a**, since the 2-position of the pyrrole ring in **1a** is quite separated from the nitro group,¹³ thus inducing the photochemical intramolecular oxygentransfer. To determine the source of the amide carbonyl oxygen of **3a**, the photoreaction of **1a** was carried out in 30% aqueous acetonitrile containing ¹⁸O-labeled water. The Mass and IR spectra of the two photoproducts obtained in this reaction showed no insertion of the labeled oxygen atom into the molecule of 2a and regioselective insertion into the carbonyl oxygen in 3a [a molecular ion peak (m/z 258) for the labeled compound $C_{10}H_6{}^{35}Cl_2N_2{}^{16}O_1{}^{18}O_1$ and a loss of $C^{18}O(m/z 30)$ from the fragment ion $[m/z 240 (M^+ - 18)]$ under ionizing radiation were observed in the mass spectrum; the stretching vibration band of the carbonyl group (v 1720 cm⁻¹ for **3a**) was significantly shifted to a lower field (ν 1691 cm⁻¹ for the ¹⁸O-labeled **3a**)]. These facts evidently indicate that the oxygen-atom at the 2-position in 3a originated from the water molecule in the media. Based on this observation and the results obtained in the tripletsensitized or intersystem-crossing accelerated experiments described above, we present a reaction sequence



Figure 6.

for the formation of **3a** in the present photoreaction of **1a** involving a single-electron oxidation of the electronrich pyrrole ring by singlet-excited nitro group ($E^{\text{red}_{1/2}}$ of **1a** = -1.31 V vs SCE in dry acetonitrile) followed by the intramolecular cyclization (cf. **B**) and the concurrent occurrence of the hydroxylation at the 2-position of the pyrrole ring by water to give the tricyclic intermediate (**C**) as shown in Scheme 2.

Analogous results were obtained for the UV irradiation of N-(4-bromobenzyl)pyrrolnitrin (1b) and the dechlorinated pyrrolnitrin $\mathbf{1c}^{21}$ in aqueous acetonitrile to afford the corresponding spiro-benzoisoxazolines (2b,c) and pyrroloindolines (3b,c), respectively. In principle, the present photoreactions seem to be general for the pyrrole antibiotics involving isopyrrolnitrin,²² oxypyrrolnitrin,²¹ dechloropyrrolnitrin,²¹ and bromonitrin.²³ 3-(3-Chloro-2nitrophenyl)-2-ethoxycarbonyl-5-methylpyrrole (5),^{21b,24} a 2,5-disubstituted analogue of 1a, was very stable even after a prolonged irradiation time (e.g., for 5 h with a 100-W UV lamp). This observation is of interest in connection with the present photoreactions and the structural design of photostable pyrrolnitrin derivatives. The isolated photoproducts **2a**-**c** and **3a**,**b** exhibited no remarkable antifungal activities against Aspergillus fumigatus, Trichophyton rubrum, and Trichophyton mentagrophytes.

III. Conclusions

The photodegradation mode of pyrrolnitrin (1a) significantly depends on the presence of water in the reaction media. In an anhydrous aprotic solvent, 1a underwent intramolecular oxidation by the nitro group on the α -position of the pyrrole ring in a triplet-excited state leading to the spiro-benzoisoxazoline 2a. In an aqueous aprotic solvent, the UV irradiation of 1a allowed the occurrence of a photochemical hydroxylation on the pyrrole ring and intramolecular cyclization with the neighboring nitro group to give the tricyclic pyrroloindoline derivative 3a. To the best of our knowledge, the photochemical hydroxylation of the pyrrole ring by water is unprecedented. These photoreactions were widely observed in the 2,5-unsubstituted pyrrolnitrins.

Experimental Section

General Procedures. All melting points were measured on Yanagimoto micro-melting point apparatus and are uncorrected. Elemental analyses were carried out in the Microana-

⁽²⁰⁾ Maki, Y.; Furuta, T.; Suzuki, M. J. Chem. Soc., Perkin Trans. *I* 1979, 553–557. Maki, Y.; Furuta, T.; Suzuki, M. Chem. Pharm. Bull., (*Tokyo*) 1979, 27, 1918–1919. Maki, Y.; Suzuki, M.; Hosogami, T.; Furuta, T. J. Chem. Soc., Perkin Trans. *I* 1974, 1354–1358. As mentioned by a reviewer, the intramolecular addition of the tripletexcited nitro group to the pyrrole ring and subsequent collapse of the adduct for the formation of 2a should involve stepwise processes, not concerted mechanisms, in response to the structural requirements for these steps.

⁽²¹⁾ a) Sako, M.; Kihara, T.; Okada, K.; Ohtani, Y.; Kawamoto, H. J. Org. Chem. **2001**, 66, 3610–3612. (b) Umio, S.; Kariyone, K.; Tanaka, K.; Kishimoto, T.; Nakamura, H.; Nishida, M. Chem. Pharm. Bull. **1970**, 18, 1414–1425. (c) Tanaka, K.; Kariyone, K.; Umio, S. Chem. Pharm. Bull. **1969**, 17, 611–615.

⁽²²⁾ Hashimoto, M.; Hattori, K. Bull. Chem. Soc. Jpn. 1966, 39, 410; Chem. Pharm. Bull. (Tokyo) 1966, 14, 1313-1316; 1968, 16, 1144.

⁽²³⁾ Ajisaka, M.; Kariyone, K.; Jomon, K. Yazawa, H.; Arima, K. J. Agric. Biochem. **1969**, *33*, 294–295.

⁽²⁴⁾ Gosteli, J. Helv. Chim. Acta **1972**, 55, 451–460. Nakano, H.; Umio, S.; Kariyone, K.; Tanaka, K.; Ueda, I.; Nakamura, H. Chem. Pharm. Bull. **1969**, 17, 567–575.

lytical Center of Gifu Pharmaceutical University. Mass spectra were measured on a JEOL JMS-SX 102A instrument with a direct inlet system operating at 70 eV. Infrared (IR) spectra were recorded on a Perkin-Elmer 1640 series FTIR spectrometer from samples as KBr pellets and ultraviolet (UV) spectra with a Shimadzu-260 spectrophotometer from samples in a dilute solution. Nuclear magnetic resonance (¹H and ¹³C NMR) spectra were obtained on a JEOL JNM-EX 400 spectrometer using deuteriochloroform or dimethyl sulfoxide as a solvent and tetramethylsilane as the internal standard. The chemical shifts are expressed in δ value (parts per million). UV Irradiation was carried out under an argon atmosphere with a Riko Rotary Photochemical Reactor (400 or 100 W highpressure mercury arc lamp; Riko Kagaku Sangyo) through a Pyrex filter at ambient temperature. A JASCO CRM-FA spectroirradiator (2 KW Xe lamp) was used for the measurement of quantum yield. Dry solvents were obtained using standard procedures. Thin-layer chromatography (TLC) analyses were performed on silica gel 60 F-254 plates (Merck Art. 5715, 0.25 mm thick) and TLC-scanning was carried out with a Shimadzu CS-9000 dual-wavelength flying-spot scanner. Rotary evaporation was carried out under reduced pressure with the bath temperature below 35 °C unless otherwise specified. Column chromatographic separation was performed with Merck silica gel 60 (70-230 mesh).

Materials. A dechlorinated pyrrolnitrin (**1c**)²¹ and 3-(3-chloro-2-nitrophenyl)-2-ethoxycarbonyl-5-methylpyrrole (**5**)^{21b,24} were prepared according to the reported procedures.

Photoreaction of Pyrrolnitrin (1a) in Anhydrous Aprotic Solvent. A solution of 1a (128 mg, 0.5 mmol) in dry acetonitrile (100 mL) was externally irradiated using a 400 W UV lamp for 10 min with stirring at ambient temperature. TLC analysis (toluene-ethyl acetate: 4/1) of the brown reaction mixture showed 35% of the starting material 1a remaining and the formation of a more polar product, together with polymeric products observed at the spotted position on the TLC plate. After removal of the solvent under reduced pressure, the resulting residual oil was subjected to a short column elution with toluene-ethyl acetate (4/1) to isolate 7,4'dichlorospiro[1,3-dihydrobenzo(c)isoxazole-3,3'-pyrrolin-2'one] (2a) (32 mg, 25%): mp 225 °C dec (a colorless amorphous powder, recrystallized from toluene); IR 3300, 3190, 1733, 1625 cm⁻¹; UV (MeCN) 286 (ϵ 3.0 \times 10³), 235 (sh, 5.0 \times 10³), 212; ¹H NMR (DMSO- d_6) 7.03 (1H, dd, J = 2, 7 Hz, H-4), 7.08 (1H, t, J = 7 Hz, H-5), 7.28 (1H, d, J = 2 Hz, H-5'), 7.41 (1H, dd, J = 2, 7 Hz, H-6), 9.96 (1H, br d, J = 2 Hz, N'H), 10.5 (1H, br, NH); ¹³C NMR (DMSO-*d*₆) 89.9, 109.7, 115.3, 120.3, 124.6, 127.5, 129.7, 130.8, 146.6, 174.8; EIMS m/z (relative intensity) 256 (M⁺ for $C_{10}H_6^{35}Cl_2N_2O_2$, 71), 228 (M⁺ – CO, 40), 221 (12), 201 (25), 193 (64). Anal. Calcd for $C_{10}H_6Cl_2N_2O_2$: C, 46.72; H, 2.35; N, 10.90. Found: C, 46.94; H, 2.50; N, 10.77. In the HMBC spectrum of 2a, peak correlations of H-4, N'H, and H-5' to $\delta_{\rm C}$ 89.9 (C-3) ppm, H-5 to $\delta_{\rm C}$ 115.3 (C-3a) ppm, H-5 to $\delta_{\rm C}$ 127.5 (C-7) ppm, H-4 to $\delta_{\rm C}$ 129.7 (C-6) ppm, H-4 and H-6 to $\delta_{\rm C}$ 146.6 (C-7a) ppm, and H-5' to $\delta_{\rm C}$ 174.8 (C-2') ppm were observed.

A solution of **1a** (2.0 mg, 7.8 μ mol) in dry THF, chloroform, or diethyl ether (2.0 mL) was irradiated in a manner similar to that described above. TLC densitometric analyses (toluene-ethyl acetate: 2/1; detector: 254 nm) of the reaction mixtures obtained after the irradiation for 10 min showed the formation of **2a** (the yields of the remaining **1a**) in 47% (15%), 60% (5%), and 72% (12%) yields, respectively. The formation of **2a** after the 20 min irradiation were 50% (in THF), 56% (in chloroform), and 76% (in diethyl ether) yields. In these reactions, the complete consumption of **1a** and the formation of polymeric polar products were observed.

Photoreactions of 1a in Aqueous Acetonitrile. A solution of **1a** (256 mg, 1.0 mmol) in 10% aqueous acetonitrile (250 mL) was externally irradiated with the 400-W UV lamp for 10 min. TLC analysis of the reaction mixture showed the formation of **2a** (17%) and a more polar product, together with the 39% recovery of **1a**. After removal of the solvent under reduced pressure, the resulting residue was subjected to a short column elution with toluene–ethyl acetate (4/1 to 2/1)

to isolate **1a** (82 mg, 32%), **2a** (38 mg, 15%), and 3,7-dichloro-8-hydroxy-8,8a-dihydropyrrolo[2,3-*b*]indol-2-one (**3a**) (85 mg, 33%): mp 175 °C dec (colorless prisms, from ethanol); IR 3255, 1720, 1665 cm⁻¹; UV (MeCN) 342 (ϵ 4.1 × 10³), 275 (sh, 4.9 × 10³), 245 (1.7 × 10⁴); ¹H NMR (DMSO-*d*₆) 5.89 (1H, d, *J* = 3 Hz, H-8a), 7.18 (1H, t, *J* = 8 Hz, H-5), 7.48 (1H, dd, *J* = 3, 8 Hz, H-6), 7.57 (1H, dd, *J* = 3, 8 Hz, H-4), 9.15 (1H, br d, *J* = 2 Hz, NH), 10.2 (1H, s, OH); ¹³C NMR (DMSO-*d*₆) 87.0, 119.5, 121.3, 122.4, 122.5, 125.4, 133.6, 146.7, 152.2, 169.8; EIMS *m/z* (relative intensity) 256 (M⁺ for C₁₀H₆³⁵Cl₂N₂O₂, 3), 238 (M⁺ - H₂O, 28), 210 (8), 66 (100). Anal. Calcd for C₁₀H₆Cl₂N₂O₂: C, 46.72; H, 2.35; N, 10.90. Found: C, 46.79; H, 2.43; N, 10.87. In the HMBC spectrum of **3a**, peak correlations of H-8a and NH to $\delta_{\rm C}$ 119.5 (C-3), H-5 to $\delta_{\rm C}$ 121.3 (C-3b), NH and H-8a to $\delta_{\rm C}$ 146.7 (C-3a), and H-4 to $\delta_{\rm C}$ 152.2 (C-7a) were observed.

After the irradiation of 1a in aqueous acetonitrile containing a variety of water-contents under analogous conditions (for 10 and 20 min), TLC densitometric analyses (toluene-ethyl acetate: 2/1; detector: 254 nm) of the reaction mixtures were carried out and showed the formation of **2a** and **3a** as follows: (a) after the 10 min irradiation: **2a** (28%) with the recovery of **1a** (35%) and no formation of **3a** in the case of anhydrous acetonitrile; 2a (24%) and 3a (20%) with a 35% recovery of 1a in the case of 5% aqueous acetonitrile; 2a (17%) and 3a (36%) with a 39% recovery of 1a in the case of 10% aqueous acetonitrile; 2a (11%) and 3a (40%) with a 44% recovery of 1a in the case of 20% aqueous acetonitrile; 2a (8%) and 3a (39%) with a 49% recovery of 1a in the case of 50% aqueous acetonitrile. (b) after the 20 min irradiation: 2a (44%) with the recovery of 1a (14%) and no formation of 3a in the case of anhydrous acetonitrile; 2a (44%) and 3a (30%) with an 18% recovery of 1a in the case of 5% aqueous acetonitrile; 2a (23%) and **3a** (47%) with a 22% recovery of **1a** in the case of 10% aqueous acetonitrile; 2a (17%) and 3a (53%) with a 26% recovery of 1a in the case of 20% aqueous acetonitrile; 2a (13%) and **3a** (48%) with a 35% recovery of **1a** in the case of 50% aqueous acetonitrile.

Quantum Yield for the Consumption of 1a in the Photolysis. The quantum yield was measured at 21 °C using potassium ferrioxalate actinometry at 280 nm. A solution of **1a** (40 mmol) in dry acetonitrile was purged well with argon. After irradiation for 6 min, the consumption of the starting **1a** was assayed by TLC densitometry eluted with toluene– ethyl acetate (2/1).

Reaction of the Photoproduct 2a with Methanol. Compound 2a (65 mg, 0.25 mmol) was dissolved in methanol (5 mL) and the solution was stirred overnight at ambient temperature. The resulting crystalline product was collected by suction to isolate 4-chloro-3-(3-chloro-2-hydroxyaminophenyl)-5-methoxy-1,5-dihydropyrrol-2-one (4) (45 mg, 62%): mp 140 °C dec; IR 3358, 3297, 1689 cm⁻¹; UV (MeCN) 295, 230 (sh), 213; ¹H NMR (DMSO-d₆) 3.20 (3H, s, OMe), 5.45 (1H, d, J = 1 Hz, H-5), 6.92 (1H, t, J = 7 Hz, H-5'), 6.95 (1H, dd, J = 2, 7 Hz, H-4'), 7.37 (1H, dd, J = 2, 7 Hz, H-6'), 7.80 (1H, br, NHOH), 8.45 (1H, br, NHOH), 8.78 (1H, br s, NH); ¹³C NMR (DMSO-d₆) 51.0, 84.9, 119, 121.2, 121.4, 129.8, 130.0, 136.4, 141.5, 145.4, 168.6; EIMS *m*/*z* (relative intensity) 288 (M⁺ for $C_{11}H_{10}{}^{35}Cl_2N_2O_3$, 17), 272 (M⁺ - O, 38), 254 (M⁺ - ${}^{35}Cl_1$, 23), 236 (22), 205 (88), 193 (100). Anal. Calcd for C₁₁H₁₀Cl₂N₂O₃: C, 45.70; H, 3.49; N, 9.69. Found: C, 45.86; H, 3.52; N, 9.66.

Effects of Triplet Sensitizer or Intersystem-crossing Accelerator on the Photoreactions of 1a in Aqueous Acetonitrile. The solution of 1a (2 mg, 7.8 μ mol) in 10% aqueous acetonitrile (2 mL) was irradiated for 5 min in the presence of acetophenone (its fluorescence was quenched by 1a in a concentration dependent manner as shown in Figure 3) or tribromomethane with a variety of concentrations and then the product distribution of 2a and 3a in these reactions were estimated by TLC densitometric analyses (toluene-ethyl acetate: 2/1; detector: 254 nm) of the reaction mixtures. The results of these reactions are shown in Figures 4 and 5.

Photoreaction of 1a in Aqueous Acetonitrile containing ¹⁸O-Labeled Water. A solution of **1a** (10 mg, 0.04 mmol) in a 30% aqueous acetonitrile containing ¹⁸O-labeled water (Euriso-top, 99.8 atm% ¹⁸O) was irradiated under the same conditions described above (for 5 min). Column chromatographic separation eluting with toluene–ethyl acetate (4/1) allowed isolation of the expected spiro-benzoisoxazoline (trace) [IR and mass spectral diversity with the unlabeled **2a** was not observed] and the ¹⁸O-labeled **3a** (2 mg, 20%) [IR 3250, 1691 cm⁻¹; EIMS *m*/*z* (relative intensity) 258 (M⁺ for C₁₀H₆³⁵-Cl₂N₂¹⁶O₁¹⁸O₁, 22), 240 (M⁺ – H₂O, 100), 223 (38), 210 (M⁺ – H₂O – C¹⁸O, 22), 206 (19), 175 (24), 148 (27)].

Preparation of 1-(4-Bromobenzyl)-3-chloro-4-(3-chloro-2-nitrophenyl)pyrrole (1b). To a solution of 1a (150 mg, 0.58 mmol) in dry N,N-dimethylformamide (50 mL) containing lithium hydride (9.3 mg, 1.17 mmol) was added 4-bromobenzyl bromide (175 mg, 0.7 mmol), and the mixture was stirred overnight at ambient temperature. After removal of the solvent under reduced pressure, the resulting residue was purified by silica gel column chromatography eluting with *n*-hexanesethyl acetate (15/1) to isolate the desired 1b (240 mg, 96%) as an oily product: ¹H NMR (CDCl₃) 4.87 (2H, s, benzylmethylene protons), 6.64 (1H, d, J = 3 Hz, H-5), 6.69 (1H, d, J = 3 Hz, H-2), 6.96 (2H, d, J = 8 Hz, H-2" and H-6"), 7.37 (2H, m, H-5" and H-6'), 7.42 (2H, d, J = 8 Hz, H-3" and H-5"), 7.49 (1H, dd, J = 3, 6 Hz, H-4'); EIMS m/z (relative intensity) 424 (M⁺ for C₁₇H₁₁⁷⁹Br³⁵Cl₂N₂O₂, 4), 169 (100). Anal. Calcd for C₁₇H₁₁-BrCl₂N₂O₂: m/z 423.9381. Found: m/z 423.9396.

Photoreaction of Pyrrolnitrin Derivatives (1b,c). A solution of 1b (240 mg, 0.56 mmol) in 50% aqueous acetonitrile (240 mL) was irradiated with the 100 W UV lamp for 2 h and the reaction mixture was evaporated to dryness. The silica gel column chromatographic separation of the resulting residue afforded the expected 1-(4-bromobenzyl)-7,4'-dichlorospiro[1,3dihydrobenzo(c)isoxazole-3,3'-pyrrolin-2'-one] (2b) (32 mg, 13%) [IR 3228, 1710 cm⁻¹; UV (MeCN) 291; ¹H NMR (DMSO-d₆) 4.54 and 4.60 (each 1H, each d, J = 16 Hz, benzylmethylene protons), 6.97 (1H, br d, J = 8 Hz, H-4), 7.05 (1H, t, J = 8 Hz, H-5), 7.24 (2H, d, J = 8 Hz, H-2" and H-6"), 7.38 (1H, s, H-5'), 7.39 (1H, br d, J = 8 Hz, H-6), 7.59 (2H, d, J = 8 Hz, H-3" and H-5"), 10.6 (1H, br, NH); ¹³C NMR (DMSO-*d*₆) 45.0, 90.3, 110.7, 115.4, 120.4, 120.9, 124.7, 127.0, 129.8, 130.0, 131.7, 133.3, 135.8, 146.5, 172.9; EIMS m/z (relative intensity) 424 (M⁺ for C₁₇H₁₁⁷⁹Br³⁵Cl₂N₂O₂, 7), 346 (8), 255 (8), 220 (8), 193 (92), 169 (100). Anal. Calcd for C₁₇H₁₁BrCl₂N₂O₂: C, 47.92; H, 2.60; N, 6,57. Found: C, 48.17; H, 2.84; N, 6.49] and 1-(4-bromobenzyl)-3,7-dichloro-8-hydroxy-8,8a-dihydropyrrolo[2,3-b]indol-2-one (3b) (97 mg, 40%) [UV (MeCN) 350, 288, 245; ¹H NMR (DMSO-d₆) 4.54 and 4.84 (each 1H, each d, J = 15 Hz, benzylmethylene protons), 5.75 (1H, s, H-8a), 7.21 (1H, t, J = 8 Hz, H-5), 7.28 (2H, d, J = 8 Hz, H-2" and H-6"), 7.49 (1H, br d, J = 8 Hz, H-6), 7.54 (2H, d, J = 8 Hz, H-3" and H-5"), 7.57 (1H, br d, J = 7 Hz, H-4), 10.4 (1H, s, NOH); ¹³C NMR (DMSO- d_6) 45.0, 89.4, 119.0, 120.5, 121.5, 122.5, 125.8, 130.2, 131.3, 131.5, 133.5, 136.1, 145.4, 152.2, 167.0; EIMS m/z (relative intensity) 424 (M⁺ for C₁₇H₁₁⁷⁹Br³⁵Cl₂N₂O₂, 10), 371 (8), 169 (100). Anal. Calcd for C₁₇H₁₁BrCl₂N₂O₂: m/z 423.9381. Found: m/z 423.9401].

Photolysis of **1c** ²¹ (90 mg, 0.4 mmol) in 50% aqueous acetonitrile (100 mL) under the same conditions followed by column chromatographic separation (toluene–ethyl acetate: 4/1) afforded 7-chloro-spiro[1,3-dihydrobenzo(*c*)isoxazole-3,3′-pyrrolin-2′-one] (**2c**) (20 mg, 22%): IR 3386, 1725 cm⁻¹; UV (MeCN) 286 (ϵ 2.0 × 10³), 245 (3.0 × 10³), 213; ¹H NMR (DMSO-*d*₆) 5.35 (1H, d, J = 5 Hz, H-4′), 6.88 (1H, br d, J = 8 Hz, H-4), 6.96 (1H, d, J = 5 Hz, H-5′), 6.98 (1H, t, J = 8 Hz, H-4), 6.96 (1H, br d, J = 8 Hz, H-6), 9.50 (1H, br, NH), 10.2 (1H, br s, N'H); ¹³C NMR (DMSO-*d*₆) 89.6, 108.4, 115.4, 120.0, 124.4, 129.0, 130.5, 134.6, 146.2, 177.4; EIMS *m/z* (relative intensity) 222 (M⁺ for C₁₀H₇³⁵Cl₁N₂O₂· 100%), 206 (M⁺ – O, 28), 204 (31), 194 (M⁺ – CO, 68), 180 (83), 178 (70). In this reaction, the expected 7-chloro-8-hydroxy-8,8a-dihydropyrolo-[2,3-*b*]indol-2-one (**3c**) was not isolated in the pure state.

Irradiation of 3-(3-Chloro-2-nitrophenyl)-2-ethoxycar-bonyl-5-methylpyrrole (5). A solution of 5 ²⁴ (10 mg, 0.03 mmol) in anhydrous acetonitrile (10 mL) was irradiated under the same conditions described above (with the 100 W UV lamp for 5h). TLC analysis of the irradiated solution showed no consumption of the starting 5.

Acknowledgment. We are grateful to the late Dr. Kazuo Kariyone of Fujisawa Pharmaceutical Co., Ltd., for his initial suggestions and continuous support of the progress in the present investigation.

Supporting Information Available: ¹H NMR, ¹³C NMR, IR, UV, and MS data for **2a**–**c**, **3a**,**b**, and **4**. Tables of crystal data, structure solution and refinement, atomic coordinates, bond lengths and angles, and anisotropic thermal parameters for **3a**. This material is available free of charge via the Internet at http://pubs.acs.org.

JO010619Z